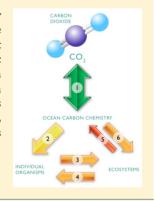


Detecting the Unexpected: A Research Framework for Ocean **Acidification**

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ABSTRACT: The threat that ocean acidification (OA) poses to marine ecosystems is now recognized and U.S. funding agencies have designated specific funding for the study of OA. We present a research framework for studying OA that describes it as a biogeochemical event that impacts individual species and ecosystems in potentially unexpected ways. We draw upon specific lessons learned about ecosystem responses from research on acid rain, carbon dioxide enrichment in terrestrial plant communities, and nitrogen deposition. We further characterize the links between carbon chemistry changes and effects on individuals and ecosystems, and enumerate key hypotheses for testing. Finally, we quantify how U.S. research funding has been distributed among these linkages, concluding that there is an urgent need for research programs designed to anticipate how the effects of OA will reverberate throughout assemblages of species.



INTRODUCTION

Rising atmospheric CO2 is linked to global warming and changes to terrestrial ecosystems. In the oceans, increasing CO2 alters surface seawater chemistry by decreasing ocean pH and calcium carbonate saturation state. 2,3 Collateral changes in biological systems are already apparent⁴ and further alterations are expected. While there is no clear boundary between the role of CO₂ in Earth's climate and its role in seawater chemistry, the term ocean acidification (OA) is used to refer to the subset of changes in ocean chemistry that propagate from the addition of anthropogenic CO₂ to seawater. On time scales shorter than centuries, the primary concern with OA is a potential reconfiguration of marine ecosystems.⁵

We suggest a research framework to guide efforts toward anticipating and projecting biological changes from OA over the coming decades to centuries. Our efforts were part of an OA Principal Investigator's Meeting sponsored by the National Science Foundation for those currently receiving OA funding from Federal funding sources within the USA. We review several past investigations into ecosystem impacts—across disparate ecosystems—and take from them lessons that we can apply to research emerging OA phenomenon. Drawing from

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these lessons, we develop a framework for integrating the effect of feedbacks between carbon chemistry changes, individual species, and ecosystems. We also use the opportunity to identify and highlight central hypotheses that need testing.

Past ecosystem-level research provides lessons that can aid future efforts to study environmental change. We look to studies of multiple ecosystems to anticipate results and formulate a conceptual framework. Specifically, we cite three case studies of the effects of environmental change on ecosystems and argue that these studies motivate a system-level approach to the study of OA.

PREVIOUS ECOLOGICAL OBSERVATIONS THAT INFORM OA RESEARCH

The first key observation from other ecological studies is that assays and experiments with single species do not necessarily predict the responses of multispecies systems. For example, an assemblage of terrestrial plants utilizes nitrogen more completely than predicted by single-species assays of nitrogen use. Thus, a diverse plant community ameliorates the effects of anthropogenic nitrogen deposition by reducing accumulation in soil. A comparison of ocean warming effects on species in situ also had a cautionary message. Along the California Current system, artificial warming of seawater by a power plant was predicted to have a positive effect on species with a southerly geographic range and a more negative effect on those with a northerly range. However, the observed species response was uncorrelated with their geographic ranges; instead, their response was suggested to be caused by species interactions.⁷ Analogously, a species that is negatively affected by OA at the individual level may ultimately benefit from OA due to indirect effects, such as decreases in the abundance of a competitor or consumer. Because studies of environmental change in the ocean will likely reveal such emergent properties—or properties that are only apparent when component parts are joined together⁸—research programs should be prepared to detect them.

A second key observation is that structural and functional properties of a system may not respond similarly. Experimental studies of acid rain into lakes revealed that the species composition of primary producers changed while their productivity (e.g., a function) remained unchanged. Thus, manipulation of a community allowed compensatory responses by some members previously at low density. We need to ask how ecosystem function changes with OA, including any direct or indirect effects on the fitness of key species.

A third key observation is that the response of an ecosystem can change through time and may include lagged responses. Terrestrial FACE (free-air CO₂ enrichment) experiments in plant communities showed an initial increase in net primary productivity as CO₂ increased, followed by an eventual decline. The lagged decline in productivity is consistent with the progressive nitrogen limitation hypothesis, where nitrogen eventually begins to limit primary productivity. Thus, only after 6 years of forest development did it become apparent that primary production would not remain elevated. 10 Trophic interactions can also take time to result in a community-level response. For example, after the elimination of three species of seed eating rodents from a desert plant community studied by Brown and Heske, 11 it took 12 years for the plant community to shift to grasses, an interval greater than predicted by the investigators. Thus, our conclusions about the effect of a perturbation may depend upon the time scales of the ecosystem

responses and the time scales of our observations of the ecosystem. Analogous to the terrestrial FACE experiments are the FOCE (free-ocean carbon dioxide enrichment) experiments (http://www.mbari.org/mars/science/foce.html), which should help reveal the temporal patterns of in situ responses of species to OA.

These three key observations are just a part of the guidance provided by previous ecological study of environmental change. As we continue our multiagency endeavors to study OA, our efforts need to (1) be informed by previous large-scale study of environmental change impacts and (2) have a framework that both guides the research and allows us to assess if we are addressing all key areas.

A RESEARCH FRAMEWORK

We propose a framework for continued study of OA that would span key questions ranging from seawater carbon chemistry to ecosystem response. We structured our framework in a triangular diagram that reflects the connectedness of different levels of study (Figure 1). First, there is still uncertainty surrounding the impacts of projected climate and ocean circulation changes on the physical and chemical processes that govern the ocean's capacity to absorb CO2, an interplay represented by the arrow labeled (1) in Figure 1. Then, changes in seawater carbon chemistry differentially affect individual organisms, populations of species, and ecosystems, often through changes to biogeochemical processes. Because effects on individual species (Arrow 2), when added together, can be nonlinear and influenced indirectly, system-level phenomena do not necessarily reflect the sum of their components (Arrows 3 and 4). Finally, we recognize that ecosystems and the carbon cycle can interact and result in an intricate feedback loop (Arrows 5 and 6).

For each labeled arrow or set of arrows, we outline the central question and key testable hypotheses (Table 1). We focus on several general and tractable hypotheses for each Arrow, and recognize that our list is not exhaustive. Arrow 1 represents a link between the atmosphere and seawater and focuses on biogeochemical hypotheses that provide the basis for more ecological hypotheses represented by Arrows 2 through 6. We also recognize that our expectations will differ in coastal versus open ocean environments. Finally, we ask how funded research projects in the U.S. cover this conceptual framework by reviewing where OA funding is being directed among these 6 arrows.

(Arrow 1) How Will the Partitioning of CO₂ between the Ocean and the Atmosphere Change with Increasing Anthropogenic CO₂? Arrow 1 is represented in green because we have a solid understanding of the basic processes responsible for the feedback between atmospheric and seawater CO_2 . The effects of anthropogenic CO_2 on ocean chemistry are relatively well understood due to research motivated—in part—by the related climate change issue of oceanic and atmospheric warming. The oceans have absorbed approximately one-third of all anthropogenic CO₂, ^{12,13} and continue to absorb approximately one-quarter of all modern emissions. 14,15 The decrease in the uptake fraction is primarily attributable to changes in ocean chemistry. Marine uptake of anthropogenic carbon increases dissolved seawater CO2 concentrations and drives the conversion of carbonate (CO_3^{2-}) to bicarbonate (HCO_3^{-}) through the net reaction:

$$CO_2 + H_2O + CO_3^{2-} \leftrightarrow 2HCO_3^{-}$$
 (1)

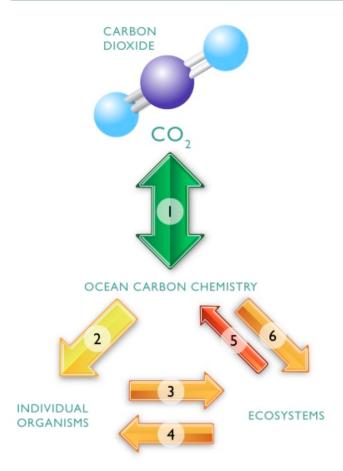


Figure 1. A framework for the study of Ocean Acidification. Each key linkage (or Arrow) is denoted with a number between 1 and 6, corresponding to a set of hypotheses presented in the text. The framework is divided into "C" for Coastal Ocean and "O" for Open Ocean. We color-coded the arrows to indicate green for a well-understood linkage and yellow for strong current research programs, while orange indicates nascent understanding in the area and red indicates a dearth of data for the linkage. We quantified the extent of current research in the U.S. associated with each arrow by tallying the studies presented in posters at the Ocean Acidification Principal Investigator's (OAPI) meeting in September 2013 (72 posters) and the U.S. National Science Foundation OA grants awarded in 2013 (n = 13), (Table 2). Our tallies slightly exceed 100% because two studies encompassed two linkages.

Another key reaction occurs increasingly as carbonate ions are depleted in seawater:

$$CO_2 + H_2O \leftrightarrow H^+ + HCO_3^-$$
 (2)

Reaction 2 increases seawater hydrogen ion concentrations and gives ocean acidification its name. Hydrographic surveys have measured changes in marine carbon chemistry resulting from CO_2 uptake with decadal resolution, half while a handful of time series have made measurements at higher frequencies. Our comparatively solid understanding of the role of CO_2 in ocean chemistry has allowed for future projections of anthropogenic changes to the oceans, although many projections focus on surface ocean pCO_2 and all rely upon uncertain predictions of future anthropogenic carbon releases.

Future research relating to Arrow 1 should address the comparatively poorly understood processes acting on \sim 100 years to millenial time scales (lagged responses), emergent

properties of the climate system that arise due to interactions between OA and other aspects of climate change, and nonlinear feedbacks between biological responses to OA and atmospheric CO_2 . As with the observations for increases in CO_2 in terrestrial systems, lagged responses are expected. Because large temporal and spatial scales involved, we suggest that hypotheses are tested using simulations from computer models constructed with the best available knowledge of OA feedbacks will be a valuable tool for Arrow 1.

Hypothesis 1a: On Time Scales of Centuries and Longer, Ocean Acidification Will Alter the Distribution of Carbon between the Atmosphere, The Upper Ocean, And the Deep Ocean. The export of biological organic matter removes carbon from the surface ocean and results in net oceanic uptake of carbon from the atmosphere, while carbonate mineral precipitation leads to a loss of carbon from the ocean to the atmosphere. If OA results in increased organic matter formation due to increased $\rm CO_2$ or inhibition of carbonate mineral cycling through decreased $\rm CO_3^{2-}$, then OA will also shift the balance between these processes and thus the partitioning of carbon between the ocean and the atmosphere. $\rm ^{29-32}$

Hypothesis 1b: Changes to Physical Processes Such As Eddy Dynamics Will Change the Partitioning of Carbon between the Ocean and Atmosphere over the Next 100 Years. Changes in atmospheric heat budgets are projected to continue to perturb wind patterns, especially over the Southern Ocean, and this may alter the natural eddy field. While the imprint of mesoscale eddies on local climate and biology has been studied,³³ it remains unclear to what degree broad scale changes in eddy dynamics (e.g., a structure) will affect natural biogeochemical cycling (e.g., a function).

Hypothesis 1c: An Intensified Hydrological Cycle Will Perturb the Historic Distribution of Freshwater Inputs to the Ocean, Thus Impacting Its Buffering Capacity. Changes in pH resulting from CO₂ uptake are buffered by seawater alkalinity. Because seawater alkalinity is linearly diluted by freshwater, and we expect regional shifts in patterns of evaporation, precipitation, sea ice cycling, riverine runoff, and glacial icemelt,³⁴ it follows that we should anticipate regional OA sensitivity changes.

Hypothesis 1d: Over Time Scales of Millennia and Longer, A Carbon Pulse from Dissolution of Sediment Minerals Will Further Alter the Chemistry of Seawater. "Carbonate compensation" refers to the dissolution of carbonate minerals stored as seafloor sediments^{35–38} and terrestrial carbonates^{39,40} in response to decreases in ocean pH. Carbonate dissolution increases seawater alkalinity, pH, and CO2 solubility, while acting as a negative feedback to changes in atmospheric CO₂. Sedimentary carbonate dissolution is proportional to both the saturation state of overlying seawater and the amount of CO₂ released within sediments through organic matter remineralization. ^{36,41} OA will lower the seawater-carbonate saturation state above sediments and has the potential to alter the ratio between organic matter and carbonate mineral export to deep sea sediments. OA will therefore likely enhance sedimentary carbonate dissolution and increase ocean alkalinity and carbon storage as a lagged ~10 000+ year response to OA.²⁷

Hypothesis 1e: The CO₂ Dynamics of an Organism's Environment Will Be Regionally Specific, And Subject to Other Anthropogenic Stressors. Recent attention has focused on the coastal oceans and marginal seas where carbon

Table 1. Summary of the Hypotheses Associated with Each Arrow Based on the Framework Provided in Figure 1, Each Hypothesis Is Discussed in Greater Detail in the Text

(C_{2})

Hypothesis 1a: On timescales of centuries and longer, ocean acidification will alter the distribution of carbon between the atmosphere and the deep ocean.

Hypothesis 1b: Changes to physical processes such as eddy dynamics will change the partitioning of carbon between ocean and the atmosphere over the next 100 years.

Hypothesis Ic: An intensified hydrological cycle and changes to freshwater inputs will impact ocean buffering capacity.

Hypothesis 1d: Over timescales of millennia and longer, a carbon pulse from dissolution of sediment minerals will further alter the chemistry of seawater.

Hypothesis 1e: The CO2 dynamics of an organism's environment will be regionally specific, and subject to other anthropogenic stressors.

Arrow 2) Are elevated CO₂ levels detrimental to marine organisms?

Hypothesis 2a: Species will differ in fitness-conferring traits affected by ocean acidification.

Hypothesis 2b: Life stages are differentially susceptible to ocean acidification.

Hypothesis 2c: Species will differ in their adaptive capacity to ocean acidification.

Hypothesis 2d: In addition to carbon chemistry changes, species will experience increases to other stressors where Hypotheses 2a, b, c above may also apply.

Arrow 3) Does OA affect organisms differentially to influence ecosystem properties?

Hypothesis 3a: If OA affects those species with strong interactions in the system, then we expect an ecosystem response in terms of function (e.g. productivity or resilience)

Hypothesis 3b: If OA affects those species with strong interactions in the system, then we expect an ecosystem response in terms of structure (e.g. taxonomic diversity)

Hypothesis 3c: If OA increases fitness of one or a few species with particular traits, then we expect an ecosystem response.

Arrow 4) Does OA change ecosystem level properties that feedback to individual species?

Hypothesis 4a: The effects of OA on a species may be mediated via biotic interactions or by abiotic conditions.

Hypothesis 4b: The effects of OA on ecosystems may establish pathways for novel indirect effects.

Arrow 5) Does ecosystem response affect carbon chemistry?

Hypothesis 5a: High primary production results in higher saturation states and decreased pCO

Hypothesis 5b: Changes in diversity and abundance of animals and phototrophs will alter carbon cycling

Hypothesis 5c: Changes in higher trophic level abundance (e.g. fishes) alter the cycling of carbon

Hypothesis 5d: Seasonality in species metabolism (e.g. remineralization) will impose higher variability in carbon cycling in coastal areas.

Arrow 6) Does OA affect ecosystem response?

Hypothesis 6a: OA will affect the structure of an ecosystem via an effect on carbon cycling

Hypothesis 6b: OA will affect the function of an ecosystem an effect on carbon cycling

chemistry can vary spatially⁴² and temporally.^{4,43} Our understanding of the expected changes to nearshore chemistry is less constrained compared to the open ocean. The added complications of this variability may lead to uncertainty in predicting the response of organisms to OA in coastal or otherwise variable regions.

In addition to the natural variability, there are other related changes to the biogeochemistry of the oceans due to climate change such as oxygen depletion, temperature increases, and changes to freshwater and nutrient inputs and cycling. These effects may be either direct or indirect and may alter the partitioning of carbon between the ocean and the atmosphere providing further feedbacks to OA. Anthropogenic nutrient loading enhances eutrophication and the same respiratory processes that lead to local hypoxia produce additional CO₂. In fact, pH changes since the onset of the industrial revolution in the eutrophication zones in the northern Gulf of Mexico are greater than would be expected solely on atmospheric CO₂. ⁴⁴

(Arrow 2) Are Elevated CO₂ Levels Detrimental to Marine Organisms? To date, the majority of effort directed at understanding the biological implications of OA has focused on individual organisms, mainly calcifying invertebrates and phototrophs (Figure 1), and there are several excellent reviews discussing this subject. 45–53 We extend this review material by focusing on four general hypotheses to enhance future research efforts.

Hypothesis 2a: Species Will Differ in Fitness-Conferring Traits Affected by Ocean Acidification. When assessing the long-term impacts of OA on marine organisms, it is important to identify the traits that most dramatically affect the fitness of a species. Fertilization success is a trait of particular importance to broadcast spawning invertebrate species⁵⁴ and shows a diversity of response to OA, including

some species with remarkable sensitivity $^{55-58}$ and others with no apparent effects. $^{59-61}$ To date, we have relatively little understanding of reproductive success in more mobile species, although elevated pCO₂ levels stimulated the reproductive output in the anemone fish, *Amphiprion melanopus*, 62 while having no impact on two copepod species. 63 Particularly for longer-lived species, however, reproductive consequences of OA over the entire life cycle remain untested and therefore unknown.

Traits that alter the cost of OA to organisms will be indirectly tied to fitness by affecting resources that could otherwise be dedicated to reproduction and survival. A nonexhaustive list includes impacts on growth, 64-66 metabolic rate, 67-69 photosynthesis, 70,71 calcification and the formation of key non-calcified materials, 53,72-76 tissue health, 77 and underlying physiological pathways. Additionally, a suite of behavioral effects has been documented with respect to olfactory and auditory sensing, 82,83 learning, 84 as well as compromised predator—prey detections, 85-88 all with serious consequences at both the individual and ecosystem level. 9 A common point between these impacts of OA is that responses are rarely uniform across organisms—even between closely related species, 90 setting the stage for shifts in relative species abundance in the oceans. Research efforts that hone in on those traits that make a species sensitive or tolerant to OA should remain a priority.

Hypothesis 2b: Life Stages Are Differentially Susceptible to Ocean Acidification. Early life stages of organisms are typically the most sensitive to environmental perturbations, which is likely also true for OA. In fact, there have been reports of reduced larval survival in fish, 91 urchins, 92 crabs, 93 and bivalves 94–96 in response to OA, although these effects are not ubiquitous. 97 Interestingly, porcelain crabs appear to be more

sensitive as juveniles than larvae, 98 emphasizing the need to understand the ontogenetic factors that define the survival thresholds for pCO₂ exposure.

Sublethal end points are also important for larval organisms in an ecological context owing to important early life environmental transitions. For example, many planktonic larvae transition to benthic habitat where they subsequently join adult populations. This recruitment period can be associated with high mortality rates and OA has been found to exacerbate this mortality in two very important ways. First, OA has been shown to inhibit olfactory capabilities of larval fish, reducing their ability to identify suitable recruitment habitat and potential predators in reef fish species. 82,85,89 These factors alone are anticipated to substantially impact the long-term health of reef fish populations, 89,99 and future work should examine impacts on other recruitment cues (e.g., auditory) and habitats (e.g., seagrass). Second, predator-based mortality during recruitment may also be further influenced by reduced growth of OA exposed individuals. It is well established that rapid growth of larval cohorts reduces vulnerability to predators by allowing individuals to more quickly reach established sizeescape thresholds. 100 By inhibiting larval growth, OA will likely extend the vulnerability window of many marine larvae. 91,99

Hypothesis 2c: Species Will Differ in Their Adaptive Capacity to Ocean Acidification. Understanding the ability of marine organisms to adapt to predicted environmental change is a crucial aspect of OA research. What species are resilient? Do species possess the capacity to adapt to a changing ocean and does this ability differ among species? When assessing these questions for OA, much can be learned from research efforts to understand global warming, which has highlighted three important areas when considering adaptive capacity: (1) evolutionary genetic change, (2) phenotypic plasticity, and (3) transgenerational plasticity.

Recent theories discussing evolutionary rescue provide guidance for the role genetic processes may play in species resilience to OA. 101,102 These theories address responses to deteriorating environments and emphasize the importance of existing genetic variation within a population that may harbor successful heritable genotypes. This may be especially true for those species living in regions of upwelling, highly variable, or otherwise extreme environments that already experience periods of high-CO₂ exposure. In addition, species with short generation times (e.g., coccolithophores) may respond to environmental alterations through adaptive evolution. 103

The importance of genetic variation in species has been demonstrated recently both with respect to genome-wide analysis ¹⁰⁴ and specific fitness-conferring traits. ^{105,106} However, our overall understanding of the scope for adaptation remains limited to relatively few species. A second important characteristic for evolutionary rescue is overall population size. Small populations are more at risk because they lack the overall scope for standing resilient genotypes. ¹⁰⁷ This is a particularly important point when considering OA and conservation strategies for harvested species. ⁵²

Phenotypic plasticity refers to the ability of organisms to alter their phenotype in response to environmental variables and thereby buffer against detrimental impacts of environmental variation. Physiological plasticity could play a key role for OA given the importance of acid—base homeostasis. ^{78,79,108} In fact, comparative physiologists have long been interested in the dynamic responses of organisms to changing environments, and much can be learned from these studies with respect to

experimental approaches. A common approach is to identify distinct populations of a species that are exposed to different environments, which has been used effectively by examining species native to naturally high-CO $_2$ areas. 104,105 A second common approach is to monitor an organism's response to environmental change through time, 78,109 often highlighting transcriptional, translational or morphological responses; however, it is important to anchor transcriptional changes to phenotype.

Transgenerational plasticity (TGP), whereby adults exposed to OA confer advantageous traits to offspring through epigenetic mechanisms, may also have implications for OA. 110,111 TGP is particularly prevalent in species coping with rapid environmental change. The current evidence for TGP suggests that some species are already adapted to rapid changes in pH and pCO2 levels, as it is a characteristic feature of many coastal habitats, and these lab studies have recently been extended to field studies demonstrating seasonal changes in pCO₂ sensitivity in Atlantic silversides (Menidia menidia). 114 Unlike the open ocean, future pCO2 levels in coastal environments may be difficult to predict; and consequently it is not known whether short-term adaptations to high CO2 via TGP may translate into long-term resilience of these species. A final consideration is the recognition that adaptive responses, via plasticity, TGP or other means, may also come with fitness costs by inducing trade-offs.

Hypothesis 2d: In Addition to Carbon Chemistry Changes, Species Will Experience Increases to Other Stressors Where Hypotheses 2a, b, c above May Also **Apply.** Ocean acidification is only one of many components of global change and, as pointed out in several reviews, 50,115 it is important that moving forward, researchers consider multiple stressors. The most obvious overlapping global climate change is the predicted increase in sea surface temperature associated with global warming, an interaction that is being increasingly explored with experiments. 116 Increasing sea surface temperatures are not only another significant stressor, but temperature has a direct effect on seawater carbonate chemistry. Important additional stressors include altered freshwater inputs into the marine environment, both through changing hydrological factors (Hypothesis 1c) and anthropogenic influences, as well as changing occurrence and duration of marine hypoxia events. Importantly, in many marine habitats hypoxia and high CO₂ occur concurrently with potential neutral, additive and/or synergistic effects of these two stressors. 117,118

Coastal ecosystems are of particular interest with respect to multistressor studies as these environments are in some cases already subject to dramatic shifts in salinity, temperature, and nutrient input, as well as daily fluctuations in oxygen and carbon dioxide. Furthermore, the coastal environment is particularly at risk from harmful anthropogenic inputs stemming from agriculture or urban runoff leading to eutrophication and polluted environments, respectively. On the one hand, because species in coastal areas typically experience high environmental variation, they may act as a reservoir for population and species resiliency; alternatively, persistent exposure to multiple stressors may increase their susceptibility to increasing OA.

As OA studies continue, it is essential that our inference is a result of from well-designed and replicated experiments. The EPOCA guide for OA addresses some essentials, ¹¹⁹ but we note that there are other important considerations. First, taxonomic representation needs to be broad. Further, the scope for

adaptation may depend upon the variability that organisms experience; natural fluctuations in pH are beginning to be included in experimental manipulations of pH. 65,117 Finally, in order to understand the functional relationship between pH changes and organism response, our experiments need to move beyond a simple treatment and control comparison to one where a number of treatment levels, including increasing levels of pCO₂ and interactions with other stressors, to understand the functional response of organisms.

(Arrow 3) Does OA's Differential Effect on Organisms Influence Ecosystem Properties? We expect differences in sensitivity among species to ocean acidification and it is these differences that will likely drive changes to the structure and function of an ecosystem. Ecosystem changes may include altered taxonomic and functional diversity, energy flow, ecosystem resilience, and structural complexity. Empirically, an investigator interested in quantifying an ecosystem response might thus measure overall carbon fixation, community calcification, nutrient sequestration or loss, or a metric of variability in multispecies abundance. Possible changes in ecosystem structure and function in response to OA are indicated by in situ studies along natural gradients in pH, pCO₂, or saturation state. Shallow, submarine CO₂ seep sites from the Mediterranean Sea, a tropical reef in Japan, and coral reefs in Papua New Guinea have provided sites to perform mensurative experiments with multiple species and functional responses, including diversity changes. Though these ecosystems are an informative addition to controlled laboratory conditions and provide a proxy for ocean acidification impacts at the ecosystem level, there are recognized limitations of present-day CO₂ seep sites. Extrapolations from seeps have limitations, because pCO2 is often more variable at seeping locations than in surrounding regions, ¹²⁰ and because seeps are comprised of open populations.

Hypothesis 3a: If OA Affects Those Species with Strong Interactions in the System, Then We Expect an Ecosystem Response in Terms of Structure (e.g., Taxonomic Diversity). Reductions in taxonomic diversity are often predicted with ocean acidification. If species losses include those species with strong interactions in the system, including keystone, engineer, or foundational species, then we expect effects to reverberate throughout the system. For example, at shallow volcanic CO₂ seeps in Papua New Guinea, pH declines from 8.1 to 7.8 without a corresponding temperature change. Increases in CO2 corresponded with reductions in hard coral species richness, including increased coverage of massive Porites and loss of diversity from structurally complex corals (e.g., branching, foliose and tabulate growth forms). ¹²¹ The implication for losses in hard and structurally complex corals could be a decline in habitat availability and quality for juvenile fish and many invertebrates. Even if the losses represent only local extirpations, local diversity reduction may still be significant. CO2 seeps at subtidal temperate sites in the Mediterranean off the coast of Ischia also exhibited reductions in taxonomic diversity with pH declines.⁵ Scleractinian corals were absent from high venting regions, and reductions in sea urchin and coralline algal abundances were significant. Following disturbance events (clearing of benthic plots) at this site, recovery patterns of rocky benthos were less variable than at ambient pH sites, resulting in homogenization and reduced functional diversity at pH levels reflective of near-future and extreme scenarios for ocean acidification. The biofilms and filamentous algae that rapidly recolonized the cleared plots at reduced pH remained abundant throughout recovery. OA effects on a species that strongly interacts in a system may in turn affect the abundance of other species, an outcome suggested in a guild of coralline algae at a site in the northeast Pacific where pH is in decline. A former competitively dominant alga has declined in thickness since the 1980s¹²⁴ and now loses approximately half of its competitive bouts, an effect that has resulted in greater intransitivity of competitive outcomes in the system. These relatively recent examples highlight in situ observations of alterations in ecosystem taxonomic and functional diversity resulting from differential sensitivity of species to reduced pH.

Hypothesis 3b: If OA Affects Those Species with Strong Interactions in the System, Then We Expect an Ecosystem Response in Terms of Function (e.g., Productivity or Resilience). Many primary producers are expected to benefit from ocean acidification, leading to an increase in ecosystem productivity. At CO₂ vents off Ischia and Papua New Guinea, seagrass cover increased with decreasing environmental pH. State Shoot densities, but not below-ground biomass, were three to four times higher at >500 ppm pCO₂. Increased seagrass and macroalgal cover corresponded with reductions in epiphytes in both vent regions.

Loss of structure-forming species, such as structurally complex corals, to ocean acidification (perhaps exacerbated by seawater temperature increases) indirectly affects macroinvertebrate groups through loss of habitat and refugia. 127 At high-CO $_2$ seep sites in Papua New Guinea, total density and representative phyla of macroinvertebrates was reduced to 48% and 85% of control values, respectively, and was associated with a reduction in reef structural complexity. Densities of those taxa that were strongly reduced at high CO $_2$ were mobile groups, while densities of sessile taxa were unaltered.

The demonstrated keystone status of calcifying species such as the seastar, *Pisaster ochraceous*, and the sea urchin, *Strongylocentrotus purpuratus*, in the northeast Pacific suggest that calcifying species such as these may initiate structural and functional changes to ecosystems. Similarly, taxa that respond to environmental change and have a central trophic position, will likely influence community structure through range changes and biotic interactions.

Hypothesis 3c: If OA Increases Fitness of One or a Few Species with Particular Traits, Then We Expect an Ecosystem Response. Even if species affected by OA are not, by themselves, strong interactors in the ecosystem, shared traits among species may lead to an ecosystem response. For example, OA may change the diversity of ecological systems, based on evidence from CO₂ vent communities where sites with decreased pH show a shift to a species-poor community dominated by fleshy algae. Mesocosm experiments where pCO₂ is manipulated also suggest a decreased diversity of macroalgae with increased pCO₂. Given the demonstrated linkages between species diversity and ecosystem function, OA can be a stressor that stimulates a negative feedback loop where diversity erodes and a functional change occurs.

(Arrow 4) Does OA Change Ecosystem Level Properties That Feedback to Individual Species? The effects of ocean acidification on individual species could also occur via indirect pathways, in which the direct effects of high CO₂ alter components of the ecosystem that inhibit those taxa not directly at risk from high CO₂. Again, many insights into indirect effects of ocean acidification are derived from in situ studies conducted along strong gradients in pCO₂, and from

mesocosm experiments that allow controlled tests of ecological interactions.

Hypothesis 4a: The Effects of OA on a Species May Be Mediated via Biotic Interactions or by Abiotic Conditions. Indirect effects of high CO₂ can be driven via biotic interactions if changes in the producer or consumer component of the ecosystem inhibit or facilitate other taxa via resource availability or competitive interaction. As an example, matforming algae in temperate kelp forests and tropical coral reefs are hypothesized to have enhanced productivity with increased CO₂ resources, ^{131,132} perhaps outcompeting kelps and corals. Hence both kelps and corals, in their respective habitats, may be indirectly inhibited by high CO₂ via space-holding competitors that are released from carbon limitation. Shifts in the competitive dominants of each respective ecosystem may result. 133 Another indirect pathway by which kelp and coral are further inhibited is by the negative effects of mat-forming algae on coral and kelp recruitment. We note, however, that if OA has deleterious effects on their grazers, algae may experience a net benefit.

Changes in resources could also lead to indirect effects on consumers via changes to the quantity and quality of their food source. 136 For example, when turf algae and their herbivores were grown in high-CO2 concentrations, the algae had increased nitrogen content. Per capita herbivory rates of gastropods, in turn, increased, suggesting elevated energetic needs under OA. 137 Similarly, negative effects of increased CO₂ on pteropods and the diverse assemblage of zooplankton may have indirect effects on the fishes that feed on them. 138 If decreases in pteropods result from their inability to calcify and maintain shells under high-CO2 conditions, then food webs in high latitude regions will be affected, as well as the substantial export flux of carbon and carbonate. A further consideration is the outcome if a species susceptible to OA is a member of a higher trophic level. The possibility of trophic cascades, or the alternate control of competitive versus consumers across all trophic levels, is a likely outcome if a top predator is affected by

Hypothesis 4b: The Effects of OA on Ecosystems May Establish Pathways for Novel Indirect Effects. Because OA likely affects biogenic structures that provide habitat to many species (e.g., corals), any dissolution of these structures or susceptibility to disturbance will indirectly affect many species. For example, the snail *Littorina littorea* fails to induce shell defenses under decreased seawater pH, a response that affects its crab predators. ¹³⁹

(Arrow 5) Does Ecosystem Response Affect Carbon Chemistry? We generally think about the effects that changing environments can have on organisms and ecosystems, but ecosystems can also have both positive and negative feedback on the environment. Changes in community diversity, structure, and abundance can all alter carbon cycling via biogeochemical reactions that impact the carbon cycle including photosynthesis, respiration, and calcification. We highlight key examples below where ecosystem responses can affect ocean carbon chemistry.

Hypothesis 5a: High Primary Production Results in Higher Saturation States and Decreased pCO₂. Primary producers utilize CO_2 through photosynthesis, which decreases pCO_2 and ultimately increases pH and $CaCO_3$ saturation state. A 10-year time series of pH data from a rocky intertidal zone in the Eastern Pacific off of Washington State showed typical diel fluctuations of about 0.24 units, reflecting net photosynthesis

during the day and net respiration at night. As subtidal macrophyte meadow in the Baltic Sea, characterized by brown algae and seagrass, showed mean diel pH variability of 0.34 units during the month of August. Additionally, a multiecosystem comparison found semidiurnal and diel pH variability patterns at different coastal locations categorized as upwelling, estuarine/near-shore, coral reef, kelp forest, and extreme. Over a 30 day period, diel pH fluctuations were 1–2 orders of magnitude larger at tropical and temperate coastal locations (0.121–1.430 units) compared to open-ocean and polar locations (0.047–0.096 units) aftect pCO_2 and pCO_3 saturation state varies regionally and has been greatest in coastal locales.

Hypothesis 5b: Changes in Diversity and Abundance of Phototrophs and Heterotrophs Will Alter Carbon **Cycling.** The diversity of both heterotrophs and phototrophs is expected to decrease as ocean pH decreases as predicted by mesocosm manipulations and studies near natural vents. 121,122,141 Loss of biodiversity could impact carbon and nutrient cycling. In grassland ecosystems, increased diversity led to increased productivity and nutrient utilization.⁶ If applied to an OA scenario, we hypothesize that decreased diversity in marine phototrophs could reduce productivity and result in incomplete nutrient utilization, leading to less fixed carbon and lower chlorophyll/nutrient ratios. Reduced primary production by phototrophs in surface waters would limit resources available for heterotrophs and would reduce carbon export out of the euphotic zone, altering global carbon cycles and nutrient utilization and potentially leading to positive feedback for the carbon content of the surface ocean.

Changes in community structure as a result of OA could impact carbon chemistry in various ways, depending on how the community structure changes. For example, in macroalgal communities, fleshy turf-forming algae may have a competitive advantage over kelp and calcareous species with increasing OA. 142,143 Different macroalgal species have different growth rates and utilize carbon and other nutrients at different rates, thereby changing carbon chemistry dynamics. With regard to heterotrophs, calcifying animals are expected to fare poorly under high $p\mathrm{CO}_2$ conditions compared to noncalcifying animals. While all animals produce CO_2 through respiration, calcifying animals also produce one mole of CO_2 for every mole of CaCO_3 produced (eq 3).

$$Ca^{2+} + 2HCO_3^- \leftrightarrow CaCO_3 + CO_2 + H_2O$$
 (3)

This production of CO_2 further reduces pH and also reduces alkalinity. Therefore, a shift away from calcifying species could impact carbon chemistry and may partially offset OA. Furthermore, in planktonic species such as pteropods, and coccolithophores CaCO_3 acts as ballast and facilitates the transport of organic carbon to the deep ocean. ^{144,145} If the abundance of such species decreases, there could be impacts on the ocean carbon pump. The abundance of heterotrophs and phototrophs is inherently linked through consumer-resource interactions such that changes in the dominant species of each group could result in imbalances. In general, if the balance shifts toward phototrophs, there could be net primary production, utilizing CO_2 and increasing pH. If the balance shifts toward animals (heterotrophs), there would be net respiration, increasing production of CO_2 and decreasing pH.

Hypothesis 5c: Changes in Higher Trophic Level Abundance (e.g., Fishes) Alter the Cycling of Carbon. Organisms occupying higher trophic levels, such as fish, also impact carbon cycling in the ocean. The importance of the role that fish play in the inorganic carbon cycle has only recently come to light. 146,147 High magnesium calcite, a form of CaCO₃ more soluble than aragonite, is produced in fish intestines as a byproduct of osmoregulation, as fish constantly drink seawater. The high-Mg calcite is excreted in mucus-coated pellets. 148 This fish intestine calcification could account for 2.7-15.4% of the estimated total new CaCO₃ production in surface oceans, ¹⁴⁶ making it a significant portion of surface ocean CaCO₃ production. The production of the CaCO3 pellets lowers alkalinity where they are produced. In shallow regions, where the pellets do not dissolve before sinking to the seafloor, their production will lower alkalinity. It has been estimated that fish contribute an average of 14% to the total estimated carbonate mud production across the Bahamian archipelago. 147 Furthermore, fish also produce H⁺ ions, which they excrete into seawater through their gills., 149 decreasing both pH and alkalinity. If fish biomass changes with OA, as indicated by overharvesting in many marine ecosystems, 130 the production of CaCO₃ and H⁺ ions by fish will also change correspondingly, altering carbon cycling.

Hypothesis 5d: Seasonality in Species Metabolism (e.g., Remineralization) Will Impose Higher Variability in Carbon Cycling in Coastal Areas. In addition to the effects of primary producers, coastal and estuarine systems are subject to carbon chemistry conditions that vary on daily and greater time scales. Additionally, coastal and estuarine systems are potentially more susceptible to OA than open oceans. 34,150,151 As these regions become eutrophic (excessive nutrient levels that lead to algal blooms), bottom waters become hypoxic or anoxic due to microbial consumption of organic matter. 152 Microbial consumption is coincident with the metabolic production of CO₂, which further reduces pH. During respiration (or remineralization), as oxygen is used up, CO₂ is produced, lowering pH. This process is generally seasonal, as the algal blooms start in spring when the water column stratifies and there is more light available, resulting in seasonally variable carbon chemistry.

(Arrow 6) Does OA Affect Ecosystem Response? Here we incorporate instances where an *ecosystem response* variable was assayed following an OA perturbation. In contrast, Arrows 2 and 3 represent the direct link between ocean carbon chemistry and an individual species (Arrow 2). A study that focused on the calcification of a particular coral species, for example, would be placed in Arrow 2, while a study that measured reef-wide calcification rate would be in Arrow 6. If OA negatively affected a single dominant species with overall fish species diversity declining, Arrow 3 would represent that process.

Increased carbon dioxide pollution, as with anthropogenic nitrogen pollution, provides a resource subsidy to some member of the system, a subsidy that can become a stress as levels increase. We thus expect changes to the carbon cycle through an increase in carbon or a relative change in the availability in the type of dissolved inorganic carbon to result in a change to the structure or function of an ecosystem.

Hypothesis 6a: OA Will Affect the Structure of an Ecosystem via an Effect on Carbon Cycling. Structural changes in an ecosystem might result from OA independent of whether single species changes have cascading or competitive effects. For example, if increased pCO₂ stimulated carbon fixation and growth of all producers relative to consumers, a structural effect on the distribution of biomass across trophic

levels would result. Similarly, structural changes are expected in ecosystems due to the negative effect of OA on calcifying species, a taxonomically diverse group. Species composition might shift while diversity does not change, as suggested by a relative dominance of picoplankton at the expense of diatoms when CO_2 and inorganic nutrients were increased in the Svalbard mesocosm experiments. ¹⁵⁴

Hypothesis 6b: OA Will Affect the Function of an Ecosystem via an Effect on Carbon Cycling. Functional ecosystem responses resulting from a direct effect of OA, presumably via the provision of greater CO₂ and HCO₃, are supported in experiments where carbon uptake and primary production increase with increasing pCO₂. 154 Although enhanced primary productivity and thus decreased pCO₂ might be thought of as a positive effect on an ecosystem service, whether nutrient limitation will inhibit the long-term carbon response, as in the Progressive Nutrient Limitation Hypothesis posited for terrestrial systems, remains to be studied. Increasing pCO₂ has been shown to increase growth of some turf algal species and phytoplankton., while kelp species did not show this response. A productivity response by phototrophs may be most likely in areas where nitrogen is increased due to anthropogenic inputs. Thus, our understanding of OA effects on ecosystem function through changes in productivity is dependent on the availability of nutrients other than carbon, as well as phototroph physiology and interactions. Combined with our discussion of Hypothesis 5a, it becomes clear that some structural and functional responses to OA will likely involve feedbacks. For example, a stimulation of primary production by increased pCO₂ could result in greater diel and seasonal variation in pH as some producers drive up pH, making carbon less accessible to other producers.

A Summary of Research Coverage to Date. Finally, we ask how ongoing research programs in the U.S. have covered this conceptual framework that spans carbon chemistry and individuals to ecosystems (Figure 1). To quantify the coverage, we used two sources of information: (1) the content of posters presented at the OA Principal Investigators Meeting in September 2013 in Washington, DC and (2) the most recent proposals funded by the NSF Ocean Acidification panel in July 2013. These two sources should integrate and be representative of the recent research efforts funded by the NSF and NOAA, as the meeting was mandatory for those receiving federal funds to do OA research and any new awardees were included in the July 2013 announcement. In total, there were 72 studies where the research content could be assigned; 15 others were excluded because they dealt with outreach or meeting summaries. We find that much of the current research is focused on understanding the interplay between the atmosphere and seawater, as well as the effects on single species (Table 2). There have also been efforts (14.3%) to understand the effects of OA on ecosystem-level responses (Arrow 6), and these have been focused in coastal systems.

We found few funded research efforts to link individual responses to system-level responses, though we acknowledge that there is ongoing research with mesocosms and at CO₂ vent sites that were not encompassed in these recent U.S. funding efforts. Thus, the scientific community has relatively little predictive power to understand how the results of scaled-down bottle and mesocosm studies will scale-up to the ecosystem level and whether emergent properties will occur (e.g., Arrow 3). Further, there was no apparent effort focusing on how system-level biological changes will feed back to ocean carbon

Table 2. We Quantified the Extent of Current Research in the US Associated with Each of the Six Main Linkages Described in Figure 1 by Tallying the Studies Presented in Posters at the Ocean Acidification Principal Investigator's (OAPI) Meeting in September 2013 (72 posters) and the U.S. National Science Foundation OA Grants Awarded in $2013 (n = 13)^a$

linkage number	linkage type	% coastal studies	% open ocean studies
1	CO ₂ to ocean carbon chemistry	14.3	11.4
2	ocean carbon chemistry to individual organisms	47.0	10.0
3	individual organisms to ecosystems	1.4	0
4	ecosystems to individual organisms	0	0
5	ecosystems to ocean carbon chemistry	1.4	0
6	ocean carbon chemistry to ecosystems	14.3	1.4

[&]quot;Our tallies slightly exceed 100% because two studies encompassed two linkages. We split these studies into groups that focused either on Coastal or Open Ocean areas.

chemistry (Arrow 5). Given that biological processes such as carbon fixation, respiration and nutrient uptake have been associated with driving substantial diel variability in coastal carbon cycles, 4,43,155 and that marine phototroph composition is predicted to change, 142,156 it is feasible that the link between system-level properties and ocean carbon (Arrow 5) could be significant.

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Notes

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REFERENCES

- (1) Menzel, A.; Sparks, T. H.; Estrella, N.; Koch, E.; Aasa, A.; Ahas, R.; Alm-Kübler, K.; Bissolli, P.; Braslavská, O.; Briede, A.; et al. European phenological response to climate change matches the warming pattern. *Global Change Biol.* **2006**, *12*, 1969–1976.
- (2) Caldeira, K.; Wickett, M. E. Oceanography: Anthropogenic carbon and ocean pH. *Nature* **2003**, *425*, 365–365.
- (3) Doney, S. C.; Fabry, V. J.; Feely, R. A.; Kleypas, J. A. Ocean acidification: The other CO₂ problem. *Ann. Rev. Mar. Sci.* **2009**, *1*, 169–192.
- (4) Wootton, J. T.; Pfister, C. A.; Forester, J. D. Dynamic patterns and ecological impacts of declining ocean pH in a high-resolution multi-year dataset. *Proc. Natl. Acad. Sci. U.S.A.* **2008**, *105*, 18848–18853.
- (5) Hall-Spencer, J. M.; Rodolfo-Metalpa, R.; Martin, S.; Ransome, E.; Fine, M.; Turner, S. M.; Rowley, S. J.; Tedesco, D.; Buia, M.-C. Volcanic carbon dioxide vents show ecosystem effects of ocean acidification. *Nature* **2008**, *454*, 96–99.

- (6) Tilman, D.; Wedin, D.; Knops, J. Productivity and sustainability influenced by biodiversity in grassland ecosystems. *Nature* **1996**, *379*, 718–720.
- (7) Schiel, D. R.; Steinbeck, J. R.; Foster, M. S. Ten years of induced ocean warming causes comprehensive changes in marine benthic communities. *Ecology* **2004**, *85*, 1833–1839.
- (8) Salt, G. W. A comment on the use of the term emergent properties. *Am. Nat.* **1979**, *113*, 145–158.
- (9) Schindler, D. W.; Mills, K. H.; Malley, D. F.; Findlay, D. L.; Shearer, J. A.; Davies, I. J.; Turner, M. A.; Linsey, G. A.; Cruikshank, D. R. Long-term ecosystem stress: The effects of years of experimental acidification on a small lake. *Science* 1985, 228, 1395–1401.
- (10) Norby, R. J.; Zak, D. R. Ecological Lessons from Free-Air CO₂ Enrichment (FACE) Experiments. *Ann. Rev. Ecol., Evol., Syst.* **2011**, 42, 181–203.
- (11) Brown, J. H.; Heske, E. J. Control of a desert-grassland transition by a keystone rodent guild. *Science* **1990**, 250, 1705–1707.
- (12) IPCC. Climate Change 2014: Impacts, Adaptation, and Vulnerability. Chapter 30; Cambridge University Press, 2014.
- (13) Sabine, C. L.; Feely, R. A. The oceanic sink for carbon dioxide. In *Greenhouse Gas Sinks*; Reay, D., Hewitt, N., Grace, J., Smith, K., Eds.; CABI Publishing: Oxfordshire, UK, 2007.
- (14) Wanninkhof, R.; Park, G.-H.; Takahashi, T.; Sweeney, C.; Feely, R.; Nojiri, Y.; Gruber, N.; Doney, S. C.; McKinley, G. A.; Lenton, A.; et al. Global ocean carbon uptake: Magnitude, variability and trends. *Biogeosciences* **2013**, *10*, 1983–2000.
- (15) Khatiwala, S.; Tanhua, T.; Mikaloff Fletcher, S.; Gerber, M.; Doney, S. C.; Graven, H. D.; Gruber, N.; McKinley, G. A.; Murata, A.; Ríos, A. F.; et al. Global ocean storage of anthropogenic carbon. *Biogeosciences* **2013**, *10*, 2169–2191.
- (16) Sabine, C. L.; Feely, R. A.; Gruber, N.; Key, R. M.; Lee, K.; Bullister, J. L.; Wanninkhof, R.; Wong, C. S.; Wallace, D. W. R.; Tilbrook, B.; et al. The oceanic sink for anthropogenic CO₂. *Science* **2004**, 305, 367–371.
- (17) Feely, R. A.; Sabine, C. L.; Hernandez-Ayon, J. M.; Ianson, D.; Hales, B. Evidence for upwelling of corrosive "acidified" water onto the continental shelf. *Science* **2008**, *320*, 1490–1492.
- (18) Byrne, R. H.; Mecking, S.; Feely, R. A.; Liu, X. Direct observations of basin-wide acidification of the North Pacific Ocean. *Geophys. Res. Lett.* **2010**, *37*, L02601.
- (19) Dore, J. E.; Lukas, R.; Sadler, D. W.; Karl, D. M. Climate-driven changes to the atmospheric CO₂ sink in the subtropical North Pacific Ocean. *Nature* **2003**, *424*, 754–757.
- (20) Wootton, J. T.; Pfister, C. A. Carbon system measurements and potential climatic drivers at a site of rapidly declining ocean pH. *PLoS One* **2012**, *7*, e53396.
- (21) Frieder, C. A.; Nam, S. H.; Martz, T. R.; Levin, L. A. High temporal and spatial variability of dissolved oxygen and pH in a nearshore California kelp forest. *Biogeosciences* **2012**, *9*, 3917–3930.
- (22) Bates, N. R.; Best, M. H. P.; Neely, K.; Garley, R.; Dickson, A. G.; Johnson, R. J. Detecting anthropogenic carbon dioxide uptake and ocean acidification in the North Atlantic Ocean. *Biogeosciences* **2012**, *9*, 2509–2522.
- (23) Bates, N.; Astor, Y.; Church, M.; Currie, K.; Dore, J.; Gonaález-Dávila, M.; Lorenzoni, L.; Muller-Karger, F.; Olafsson, J.; Santa-Casiano, M. A Time-Series View of Changing Ocean Chemistry due to ocean uptake of anthropogenic CO₂ and ocean acidification. *Oceanography* **2014**, *27*, 126–141.
- (24) Caldeira, K.; Wickett, M. E. Ocean model predictions of chemistry changes from carbon dioxide emissions to the atmosphere and ocean. *J. Geophys. Res.* **2005**, *110*, C09S04.
- (25) Feely, R.; Doney, S.; Cooley, S. Ocean acidification: Present conditions and future changes in a high-CO₂ world. *Oceanography* **2009**, 22, 36–47.
- (26) Dunne, J. P.; John, J. G.; Adcroft, A. J.; Griffies, S. M.; Hallberg, R. W.; Shevliakova, E.; Stouffer, R. J.; Cooke, W.; Dunne, K. A.; Harrison, M. J.; et al. GFDL's ESM2 global coupled climate—carbon earth system models. Part I: Physical formulation and baseline simulation characteristics. *J. Clim.* 2012, 25, 6646—6665.

- (27) Zeebe, R. E. History of seawater carbonate chemistry, atmospheric CO₂, and ocean acidification. *Ann. Rev. Earth Planet. Sci.* **2012**, *40*, 141–165.
- (28) Volk, T.; Hoffert, M. I. The Carbon Cycle and Atmospheric CO₂: Natural Variations Archean to Present; American Geophysical Union, 1985.
- (29) Ito, T.; Follows, M. J. Preformed phosphate, soft tissue pump and atmospheric CO₂. J. Mar. Res. 2005, 63, 813-839.
- (30) Marinov, I.; Gnanadesikan, A.; Sarmiento, J. L.; Toggweiler, J. R.; Follows, M.; Mignone, B. K. Impact of oceanic circulation on biological carbon storage in the ocean and atmospheric pCO₂. *Global Biogeochem. Cycles* **2008**, 22, GB3007.
- (31) Goodwin, P.; Williams, R. G.; Ridgwell, A.; Follows, M. J. Climate sensitivity to the carbon cycle modulated by past and future changes in ocean chemistry. *Nat. Geosci* **2009**, *2*, 145–150.
- (32) Kwon, E. Y.; Sarmiento, J. L.; Toggweiler, J. R.; DeVries, T. The control of atmospheric *p*CO₂ by ocean ventilation change: The effect of the oceanic storage of biogenic carbon: Ventilation control of atmospheric CO₂. *Global Biogeochem. Cycles* **2011**, *25*, GB3026.
- (33) Frenger, I.; Gruber, N.; Knutti, R.; Münnich, M. Imprint of southern ocean eddies on winds, clouds and rainfall. *Nat. Geosci* **2013**, *6*, 608–612.
- (34) Durack, P. J.; Wijffels, S. E.; Matear, R. J. Ocean salinities reveal strong global water cycle intensification during 1950 to 2000. *Science* **2012**, 336, 455–458.
- (35) Broecker, W. S.; Peng, T.-H. The role of CaCO₃ compensation in the glacial to interglacial atmospheric CO₂ change. *Global Biogeochem. Cycles* **1987**, *1*, 15–29.
- (36) Hales, B. Respiration, dissolution, and the lysocline. *Paleoceanography* **2003**, *18*, 1099.
- (37) Archer, D. Fate of fossil fuel CO₂ in geologic time. *J. Geophys. Res.* **2005**, *110*, C09S05.
- (38) Eby, M.; Zickfeld, K.; Montenegro, A.; Archer, D.; Meissner, K. J.; Weaver, A. J. Lifetime of anthropogenic climate change: Millennial time scales of potential CO_2 and surface temperature perturbations. *J. Clim.* **2009**, 22, 2501–2511.
- (39) Raymond, P. A.; Cole, J. J. Increase in the export of alkalinity from North America's largest river. *Science* **2003**, *301*, 88–91.
- (40) Kaushal, S. S.; Likens, G. E.; Utz, R. M.; Pace, M. L.; Grese, M.; Yepsen, M. Increased river alkalinization in the Eastern U.S. *Environ. Sci. Technol.* **2013**, 47, 10302–10311.
- (41) Hales, B.; Emerson, S. Calcite dissolution in sediments of the Ontong-Java Plateau: In situ measurements of pore water O₂ and pH. *Global Biogeochem. Cycles* **1996**, *10*, 527–541.
- (42) Hofmann, G. E.; Smith, J. E.; Johnson, K. S.; Send, U.; Levin, L. A.; Micheli, F.; Paytan, A.; Price, N. N.; Peterson, B.; Takeshita, Y.; et al. High-frequency dynamics of ocean pH: A multi-ecosystem comparison. *PLoS One* **2011**, *6*, e28983.
- (43) Baumann, H.; Wallace, R. B.; Tagliaferri, T.; Gobler, C. J. Large natural pH, CO₂ and O₂ fluctuations in a temperate tidal salt marsh on diel, seasonal, and interannual time scales. *Estuaries Coasts* **2014**, 1–12.
- (44) Cai, W.-J.; Hu, X.; Huang, W.-J.; Murrell, M. C.; Lehrter, J. C.; Lohrenz, S. E.; Chou, W.-C.; Zhai, W.; Hollibaugh, J. T.; Wang, Y.; et al. Acidification of subsurface coastal waters enhanced by eutrophication. *Nat. Geosci* **2011**, *4*, 766–770.
- (45) Langer, G. N. G. Strain-specific responses of *Emiliania huxleyi* to changing seawater carbonate chemistry. *Biogeosciences* **2009**, *6*, 2637–2646.
- (46) Dupont, S.; Ortega-Martínez, O.; Thorndyke, M. Impact of near-future ocean acidification on echinoderms. *Ecotoxicology* **2010**, 19, 449–462.
- (47) Hofmann, G. E.; Todgham, A. E. Living in the now: Physiological mechanisms to tolerate a rapidly changing environment. *Annu. Rev. Physiol.* **2010**, 72, 127–145.
- (48) Pörtner, H. O.; Peck, M. A. Climate change effects on fishes and fisheries: Towards a cause-and-effect understanding. *J. Fish Biol.* **2010**, 77, 1745–1779.

- (49) Munday, P. L.; McCormick, M. I.; Nilsson, G. E. Impact of global warming and rising CO₂ levels on coral reef fishes: What hope for the future? *J. Exp. Biol.* **2012**, *215*, 3865–3873.
- (50) Bijma, J.; Pörtner, H.-O.; Yesson, C.; Rogers, A. D. Climate change and the oceans—What does the future hold? *Mar. Pollut. Bull.* **2013**, *74*, 495–505.
- (51) Kelly, M. W.; Hofmann, G. E. Adaptation and the physiology of ocean acidification. *Funct. Ecol* **2013**, *27*, 980–990.
- (52) Munday, P. L.; Warner, R. R.; Monro, K.; Pandolfi, J. M.; Marshall, D. J. Predicting evolutionary responses to climate change in the sea. *Ecol. Lett.* **2013**, *16*, 1488–1500.
- (53) Van de Waal, D. B.; John, U.; Ziveri, P.; Reichart, G.-J.; Hoins, M.; Sluijs, A.; Rost, B. Ocean acidification reduces growth and calcification in a marine dinoflagellate. *PLoS One* **2013**, *8*, e65987.
- (54) Byrne, M. Global change ecotoxicology: Identification of early life history bottlenecks in marine invertebrates, variable species responses and variable experimental approaches. *Mar. Environ. Res.* **2012**, *76*, 3–15.
- (55) Havenhand, J. N.; Buttler, F.-R.; Thorndyke, M. C.; Williamson, J. E. Near-future levels of ocean acidification reduce fertilization success in a sea urchin. *Curr. Biol.* **2008**, *18*, R651–R652.
- (56) Albright, R.; Mason, B.; Miller, M.; Langdon, C. Ocean acidification compromises recruitment success of the threatened Caribbean coral *Acropora palmata*. *Proc. Natl. Acad. Sci. U.S.A.* **2010**, 107, 20400–20404.
- (57) Albright, R.; Mason, B. Projected near-future levels of temperature and pCO₂ reduce coral fertilization success. *PLoS One* **2013**, *8*, e56468.
- (58) Frieder, C. A. Present-day nearshore pH differentially depresses fertilization in congeneric sea urchins. *Biol. Bull.* **2014**, 226, 1–7.
- (59) Byrne, M.; Ho, M.; Selvakumaraswamy, P.; Nguyen, H. D.; Dworjanyn, S. A.; Davis, A. R. Temperature, but not pH, compromises sea urchin fertilization and early development under near-future climate change scenarios. *Proc. Biol. Sci.* **2009**, *276*, 1883–1888.
- (60) Byrne, M.; Soars, N.; Selvakumaraswamy, P.; Dworjanyn, S. A.; Davis, A. R. Sea urchin fertilization in a warm, acidified and high pCO₂ ocean across a range of sperm densities. *Mar. Environ. Res.* **2010**, *69*, 234–239.
- (61) Ho, M. A.; Price, C.; King, C. K.; Virtue, P.; Byrne, M. Effects of ocean warming and acidification on fertilization in the Antarctic echinoid *Sterechinus neumayeri* across a range of sperm concentrations. *Mar. Environ. Res.* **2013**, *90*, 136–141.
- (62) Miller, G. M.; Watson, S.-A.; Donelson, J. M.; McCormick, M. I.; Munday, P. L. Parental environment mediates impacts of increased carbon dioxide on a coral reef fish. *Nat. Clim. Change* **2012**, *2*, 858–861
- (63) McConville, K.; Halsband, C.; Fileman, E. S.; Somerfield, P. J.; Findlay, H. S.; Spicer, J. I. Effects of elevated CO_2 on the reproduction of two calanoid copepods. *Mar. Pollut. Bull.* **2013**, 73, 428–434.
- (64) Moran, D.; Støttrup, J. G. The effect of carbon dioxide on growth of juvenile Atlantic cod *Gadus morhua L. Aquat. Toxicol.* **2011**, 102, 24–30.
- (65) Bramanti, L.; Movilla, J.; Guron, M.; Calvo, E.; Gori, A.; Dominguez-Carrió, C.; Grinyó, J.; Lopez-Sanz, A.; Martinez-Quintana, A.; Pelejero, C.; et al. Detrimental effects of ocean acidification on the economically important Mediterranean red coral (*Corallium rubrum*). *Glob. Change Biol.* **2013**, *19*, 1897–1908.
- (66) Cornwall, C. E.; Hepburn, C. D.; McGraw, C. M.; Currie, K. I.; Pilditch, C. A.; Hunter, K. A.; Boyd, P. W.; Hurd, C. L. Diurnal fluctuations in seawater pH influence the response of a calcifying macroalga to ocean acidification. *Proc. Biol. Sci.* **2013**, 280, 20132201.
- (67) Stumpp, M.; Dupont, S.; Thorndyke, M. C.; Melzner, F. $\rm CO_2$ induced seawater acidification impacts sea urchin larval development II: Gene expression patterns in pluteus larvae. *Comp. Biochem. Physiol.*, *Part A Mol. Integr. Physiol.* **2011**, *160*, 320–330.
- (68) Carter, H. A.; Ceballos-Osuna, L.; Miller, N. A.; Stillman, J. H. Impact of ocean acidification on metabolism and energetics during early life stages of the intertidal porcelain crab *Petrolisthes cinctipes. J. Exp. Biol.* **2013**, 216, 1412–1422.

- (69) Enzor, L. A.; Zippay, M. L.; Place, S. P. High latitude fish in a high ${\rm CO_2}$ world: Synergistic effects of elevated temperature and carbon dioxide on the metabolic rates of Antarctic notothenioids. Comp. Biochem. Physiol., Part A Mol. Integr. Physiol. **2013**, 164, 154–161.
- (70) Xu, J.; Gao, K. Future CO₂-Induced ocean acidification mediates the physiological performance of a green tide alga1. *Plant Physiol.* **2012**, *160*, 1762–1769.
- (71) Yang, G.; Gao, K. Physiological responses of the marine diatom *Thalassiosira pseudonana* to increased pCO_2 and seawater acidity. *Mar. Environ. Res.* **2012**, *79*, 142–151.
- (72) Byrne, M.; Ho, M. A.; Koleits, L.; Price, C.; King, C. K.; Virtue, P.; Tilbrook, B.; Lamare, M. Vulnerability of the calcifying larval stage of the Antarctic sea urchin *Sterechinus neumayeri* to near-future ocean acidification and warming. *Glob. Change Biol.* **2013**, *19*, 2264–2275.
- (73) Byrne, M.; Lamare, M.; Winter, D.; Dworjanyn, S. A.; Uthicke, S. The stunting effect of a high CO₂ ocean on calcification and development in sea urchin larvae, a synthesis from the tropics to the poles. *Philos. Trans. R. Soc. B* **2013**, 368, 20120439.
- (74) Crook, E. D.; Cohen, A. L.; Rebolledo-Vieyra, M.; Hernandez, L.; Paytan, A. Reduced calcification and lack of acclimatization by coral colonies growing in areas of persistent natural acidification. *Proc. Natl. Acad. Sci. U.S.A.* **2013**, 11044–11049.
- (75) Kato, A.; Hikami, M.; Kumagai, N. H.; Suzuki, A.; Nojiri, Y.; Sakai, K. Negative effects of ocean acidification on two crustose coralline species using genetically homogeneous samples. *Mar. Environ. Res.* **2014**, *94*, 1–6.
- (76) O'Donnell, M. J.; George, M. N.; Carrington, E. Mussel byssus attachment weakened by ocean acidification. *Nat. Clim. Change* **2013**, 3, 587–590.
- (77) Frommel, A. Y.; Maneja, R.; Lowe, D.; Malzahn, A. M.; Geffen, A. J.; Folkvord, A.; Piatkowski, U.; Reusch, T. B. H.; Clemmesen, C. Severe tissue damage in Atlantic cod larvae under increasing ocean acidification. *Nat. Clim. Change* **2012**, *2*, 42–46.
- (78) Esbaugh, A. J.; Heuer, R.; Grosell, M. Impacts of ocean acidification on respiratory gas exchange and acid-base balance in a marine teleost, *Opsanus beta. J. Comp. Physiol., B* **2012**, *182*, 921–934.
- (79) Heuer, R. M.; Esbaugh, A. J.; Grosell, M. Ocean acidification leads to counterproductive intestinal base loss in the gulf toadfish (*Opsanus beta*). *Physiol. Biochem. Zool.* **2012**, *85*, 450–459.
- (80) Nilsson, G. E.; Dixson, D. L.; Domenici, P.; McCormick, M. I.; Sørensen, C.; Watson, S.-A.; Munday, P. L. Near-future carbon dioxide levels alter fish behaviour by interfering with neurotransmitter function. *Nat. Clim. Change* **2012**, *2*, 201–204.
- (81) Strobel, A.; Bennecke, S.; Leo, E.; Mintenbeck, K.; Pörtner, H. O.; Mark, F. C. Metabolic shifts in the Antarctic fish *Notothenia rossii* in response to rising temperature and pCO₂. *Front. Zool.* **2012**, *9*, 28.
- (82) Munday, P. L.; Dixson, D. L.; Donelson, J. M.; Jones, G. P.; Pratchett, M. S.; Devitsina, G. V.; Døving, K. B. Ocean acidification impairs olfactory discrimination and homing ability of a marine fish. *Proc. Natl. Acad. Sci. U.S.A.* **2009**, *106*, 1848–1852.
- (83) Simpson, S. D.; Munday, P. L.; Wittenrich, M. L.; Manassa, R.; Dixson, D. L.; Gagliano, M.; Yan, H. Y. Ocean acidification erodes crucial auditory behaviour in a marine fish. *Biol. Lett.* **2011**, *7*, 917–920.
- (84) Ferrari, M. C. O.; Manassa, R. P.; Dixson, D. L.; Munday, P. L.; McCormick, M. I.; Meekan, M. G.; Sih, A.; Chivers, D. P. Effects of ocean acidification on learning in coral reef fishes. *PLoS One* **2012**, *7*, e31478.
- (85) Dixson, D. L.; Munday, P. L.; Jones, G. P. Ocean acidification disrupts the innate ability of fish to detect predator olfactory cues. *Ecol. Lett.* **2010**, *13*, 68–75.
- (86) Cripps, I. L.; Munday, P. L.; McCormick, M. I. Ocean acidification affects prey detection by a predatory reef fish. *PLoS One* **2011**, *6*, e22736.
- (87) Ferrari, M. C. O.; Dixson, D. L.; Munday, P. L.; McCormick, M. I.; Meekan, M. G.; Sih, A.; Chivers, D. P. Intrageneric variation in antipredator responses of coral reef fishes affected by ocean

- acidification: Implications for climate change projections on marine communities. *Global Change Biol.* **2011**, *17*, 2980–2986.
- (88) Ferrari, M. C. O.; McCormick, M. I.; Munday, P. L.; Meekan, M. G.; Dixson, D. L.; Lonnstedt, Ö.; Chivers, D. P. Putting prey and predator into the CO₂ equation—Qualitative and quantitative effects of ocean acidification on predator-prey interactions. *Ecol. Lett.* **2011**, *14*, 1143–1148.
- (89) Munday, P. L.; Dixson, D. L.; McCormick, M. I.; Meekan, M.; Ferrari, M. C. O.; Chivers, D. P. Replenishment of fish populations is threatened by ocean acidification. *Proc. Natl. Acad. Sci. U.S.A.* **2010**, 107, 12930–12934.
- (90) Munday, P. L.; Gagliano, M.; Donelson, J. M.; Dixson, D. L.; Thorrold, S. R. Ocean acidification does not affect the early life history development of a tropical marine fish. *Mar. Ecol.: Prog. Ser.* **2011**, 423, 211–221.
- (91) Baumann, H.; Talmage, S. C.; Gobler, C. J. Reduced early life growth and survival in a fish in direct response to increased carbon dioxide. *Nat. Clim. Change* **2012**, *2*, 38–41.
- (92) Dorey, N.; Lançon, P.; Thorndyke, M.; Dupont, S. Assessing physiological tipping point of sea urchin larvae exposed to a broad range of pH. *Global Change Biol.* **2013**, *19*, 3355–3367.
- (93) Long, W. C.; Swiney, K. M.; Harris, C.; Page, H. N.; Foy, R. J. Effects of ocean acidification on Juvenile Red King Crab (*Paralithodes camtschaticus*) and Tanner Crab (*Chionoecetes bairdi*) growth, condition, calcification, and survival. *PLoS One* **2013**, *8*, e60959.
- (94) Talmage, S. C.; Gobler, C. J. Effects of past, present, and future ocean carbon dioxide concentrations on the growth and survival of larval shellfish. *Proc. Natl. Acad. Sci. U.S.A.* **2010**, *107*, 17246–17251.
- (95) Dickinson, G. H.; Ivanina, A. V.; Matoo, O. B.; Pörtner, H. O.; Lannig, G.; Bock, C.; Beniash, E.; Sokolova, I. M. Interactive effects of salinity and elevated CO₂ levels on juvenile eastern oysters *Cassostrea virginica*. *J. Exp. Biol.* **2012**, 215, 29–43.
- (96) Van Colen, C.; Debusschere, E.; Braeckman, U.; Van Gansbeke, D.; Vincx, M. The early life history of the clam *Macoma balthica* in a high CO₂ World. *PLoS One* **2012**, 7, e44655.
- (97) Hurst, T. P.; Fernandez, E. R.; Mathis, J. T.; Miller, J. A.; Stinson, C. M.; Ahgeak, E. F. Resiliency of juvenile walleye pollock to projected levels of ocean acidification. *Aquat. Biol.* **2012**, *17*, 247–259.
- (98) Ceballos-Osuna, L.; Carter, H. A.; Miller, N. A.; Stillman, J. H. Effects of ocean acidification on early life-history stages of the intertidal porcelain crab *Petrolisthes cinctipes. J. Exper. Biol.* **2013**, *216*, 1405–1411.
- (99) Doropoulos, C.; Ward, S.; Marshell, A.; Diaz-Pulido, G.; Mumby, P. J. Interactions among chronic and acute impacts on coral recruits: The importance of size-escape thresholds. *Ecology* **2012**, *93*, 2131–2138.
- (100) Crowder, L. B.; Rice, J. A.; Miller, T. J.; Marschall, E. A. Empirical and theoretical approaches to size-based interactions and recruitment variability in fishes. In *Individual-Based Models and Approaches in Ecology*; DeAngelis, D. L., Gross, L. J., Eds.; Springer, 1992; pp 237–255.
- (101) Bell, G. Evolutionary rescue and the limits of adaptation. *Philos. Trans. R. Soc. B* **2013**, *368*, 20120080.
- (102) Gonzalez, A.; Ronce, O.; Ferriere, R.; Hochberg, M. E. Evolutionary rescue: An emerging focus at the intersection between ecology and evolution. *Philos. Trans. R. Soc. B* **2013**, *368*, 20120404.
- (103) Lohbeck, K. T.; Riebesell, U.; Reusch, T. B. H. Adaptive evolution of a key phytoplankton species to ocean acidification. *Nat. Geosci.* **2012**, *5*, 346–351.
- (104) Pespeni, M. H.; Sanford, E.; Gaylord, B.; Hill, T. M.; Hosfelt, J. D.; Jaris, H. K.; LaVigne, M.; Lenz, E. A.; Russell, A. D.; Young, M. K.; et al. Evolutionary change during experimental ocean acidification. *Proc. Natl. Acad. Sci. U.S.A.* **2013**, *110*, 6937–6942.
- (105) Calosi, P.; Rastrick, S. P. S.; Lombardi, C.; de Guzman, H. J.; Davidson, L.; Jahnke, M.; Giangrande, A.; Hardege, J. D.; Schulze, A.; Spicer, J. I.; et al. Adaptation and acclimatization to ocean acidification in marine ectotherms: An in situ transplant experiment with polychaetes at a shallow CO_2 vent system. *Philos. Trans. R. Soc., B* **2013**, 368, 20120444.

- (106) Kelly, M. W.; Padilla-Gamiño, J. L.; Hofmann, G. E. Natural variation and the capacity to adapt to ocean acidification in the keystone sea urchin *Strongylocentrotus purpuratus*. *Global Change Biol.* **2013**, *19*, 2536–2546.
- (107) Wootton, J. T.; Pfister, C. A. Experimental separation of genetic and demographic factors on extinction risk in wild populations. *Ecology* **2013**, *94*, 2117–2123.
- (108) Stumpp, M.; Hu, M. Y.; Melzner, F.; Gutowska, M. A.; Dorey, N.; Himmerkus, N.; Holtmann, W. C.; Dupont, S. T.; Thorndyke, M. C.; Bleich, M. Acidified seawater impacts sea urchin larvae pH regulatory systems relevant for calcification. *Proc. Natl. Acad. Sci. U.S.A.* **2012**, *109*, 18192–18197.
- (109) Todgham, A. E.; Hofmann, G. E. Transcriptomic response of sea urchin larvae *Strongylocentrotus purpuratus* to CO₂-driven seawater acidification. *I. Exv. Biol.* **2009**, *212*, 2579–2594.
- (110) Miller, G. M.; Watson, S.-A.; Donelson, J. M.; McCormick, M. L.; Munday, P. L. Parental environment mediates impacts of increased carbon dioxide on a coral reef fish. *Nat. Clim. Change* **2012**, *2*, 858–861
- (111) Parker, L. M.; Ross, P. M.; O'Connor, W. A.; Borysko, L.; Raftos, D. A.; Pörtner, H.-O. Adult exposure influences offspring response to ocean acidification in oysters. *Global Change Biol.* **2012**, *18*, 82–92.
- (112) Salinas, S.; Munch, S. B. Thermal legacies: Transgenerational effects of temperature on growth in a vertebrate. *Ecol. Lett.* **2012**, *15*, 159–163.
- (113) Salinas, S.; Brown, S. C.; Mangel, M.; Munch, S. B. Nongenetic inheritance and changing environments. *Non-Genet. Inheritance* **2013**. *1*. 38–50.
- (114) Murray, C. M.; Malvezzi, A.; Gobler, C. J.; Baumann, H. Offspring sensitivity to ocean acidification changes seasonally in a coastal marine fish. *Mar. Ecol.: Prog. Ser.* **2014**, *504*, 1–11.
- (115) Dupont, S.; Pörtner, H. Marine science: Get ready for ocean acidification. *Nature* **2013**, *498*, 429–429.
- (116) Rodolfo-Metalpa, R.; Houlbrèque, F.; Tambutté, É.; Boisson, F.; Baggini, C.; Patti, F. P.; Jeffree, R.; Fine, M.; Foggo, A.; Gattuso, J.-P.; et al. Coral and mollusc resistance to ocean acidification adversely affected by warming. *Nat. Clim. Change* **2011**, *1*, 308–312.
- (117) Frieder, C. A.; Gonzalez, J. P.; Bockmon, E. E.; Navarro, M. O.; Levin, L. A. Can variable pH and low oxygen moderate ocean acidification outcomes for mussel larvae? *Global Chang Biol.* **2014**, 20, 754–764.
- (118) Gobler, C. J.; DePasquale, E. L.; Griffith, A. W.; Baumann, H. Hypoxia and acidification have additive and synergistic negative effects on the growth, survival, and metamorphosis of early life stage bivalves. *PLoS One* **2014**, *9*, e83648.
- (119) Guide to Best Practices for Ocean Acidification Research and Data Reporting; Riebesell, U., Fabry, V. J., Hansson, L., Gattuso, J.-P., Eds.; Publications Office of the European Union: Luxembourg, 2010.
- (120) Kerrison, P.; Hall-Spencer, J. M.; Suggett, D. J.; Hepburn, L. J.; Steinke, M. Assessment of pH variability at a coastal CO₂ vent for ocean acidification studies. *Estuarine, Coastal Shelf Sci.* **2011**, 94, 129–137.
- (121) Fabricius, K. E.; Langdon, C.; Uthicke, S.; Humphrey, C.; Noonan, S.; De'ath, G.; Okazaki, R.; Muehllehner, N.; Glas, M. S.; Lough, J. M. Losers and winners in coral reefs acclimatized to elevated carbon dioxide concentrations. *Nat. Clim. Change* **2011**, *1*, 165–169.
- (122) Kroeker, K. J.; Gambi, M. C.; Micheli, F. Community dynamics and ecosystem simplification in a high-CO₂ ocean. *Proc. Natl. Acad. Sci. U. S. A.* **2013**, *110*, 12721–12726.
- (123) Milazzo, M.; Rodolfo-Metalpa, R.; Chan, V. B. S.; Fine, M.; Alessi, C.; Thiyagarajan, V.; Hall-Spencer, J. M.; Chemello, R. Ocean acidification impairs vermetid reef recruitment. *Sci. Rep.* **2014**, *4*.
- (124) McCoy, S. J. Morphology of the crustose coralline alga *Pseudolithophyllum muricatum* (Corallinales, Rhodophyta) responds to 30 years of ocean acidification in the Northeast Pacific. *J. Phycol.* **2013**, 49, 830–837.

- (125) McCoy, S. J.; Pfister, C. A. Historical comparisons reveal altered competitive interactions in a guild of crustose coralline algae. *Ecol. Lett.* **2014**, *17*, 475–483.
- (126) Martin, S.; Rodolfo-Metalpa, R.; Ransome, E.; Rowley, S.; Buia, M.-C.; Gattuso, J.-P.; Hall-Spencer, J. Effects of naturally acidified seawater on seagrass calcareous epibionts. *Biol. Lett.* **2008**, *4*, 689–692.
- (127) Fabricius, K. E.; De'ath, G.; Noonan, S.; Uthicke, S. Ecological effects of ocean acidification and habitat complexity on reef-associated macroinvertebrate communities. *Proc. R. Soc. B* **2014**, *281*, 20132479.
- (128) Kroeker, K. J.; Micheli, F.; Gambi, M. C.; Martz, T. R. Divergent ecosystem responses within a benthic marine community to ocean acidification. *Proc. Natl. Acad. Sci. U.S.A.* **2011**, *108*, 14515–14520.
- (129) Asnaghi, V.; Chiantore, M.; Mangialajo, L.; Gazeau, F.; Francour, P.; Alliouane, S.; Gattuso, J.-P. Cascading effects of ocean acidification in a rocky subtidal community. *PLoS One* **2013**, *8*, e61978.
- (130) Worm, B.; Barbier, E. B.; Beaumont, N.; Duffy, J. E.; Folke, C.; Halpern, B. S.; Jackson, J. B. C.; Lotze, H. K.; Micheli, F.; Palumbi, S. R.; et al. Impacts of biodiversity loss on ocean ecosystem services. *Science* **2006**, *314*, 787–790.
- (131) Connell, S. D.; Russell, B. D. The direct effects of increasing ${\rm CO_2}$ and temperature on non-calcifying organisms: Increasing the potential for phase shifts in kelp forests. *Proc. R. Soc. B* **2010**, 277, 1409–1415.
- (132) Hepburn, C. D.; Pritchard, D. W.; Cornwall, C. E.; McLeod, R. J.; Beardall, J.; Raven, J. A.; Hurd, C. L. Diversity of carbon use strategies in a kelp forest community: Implications for a high $\rm CO_2$ ocean. *Global Change Biol.* **2011**, *17*, 2488–2497.
- (133) Connell, S. D.; Kroeker, K. J.; Fabricius, K. E.; Kline, D. I.; Russell, B. D. The other ocean acidification problem: CO_2 as a resource among competitors for ecosystem dominance. *Philos. Trans. R. Soc. B* **2013**, 368, 20120442.
- (134) Gorman, D.; Connell, S. D. Recovering subtidal forests in human-dominated landscapes. *J. Appl. Ecol.* **2009**, *46*, 1258–1265.
- (135) Vermeij, M. J. A.; van Moorselaar, I.; Engelhard, S.; Hörnlein, C.; Vonk, S. M.; Visser, P. M. The effects of nutrient enrichment and herbivore abundance on the ability of turf algae to overgrow coral in the Caribbean. *PLoS One* **2010**, *5*, e14312.
- (136) Andersen, T.; Elser, J. J.; Hessen, D. O. Stoichiometry and population dynamics. *Ecol. Lett.* **2004**, *7*, 884–900.
- (137) Falkenberg, L. J.; Russell, B. D.; Connell, S. D. Future herbivory: The indirect effects of enriched CO₂ may rival its direct effects. *Mar. Ecol.: Prog. Ser.* **2013**, 492, 85–95.
- (138) Orr, J. C.; Fabry, V. J.; Aumont, O.; Bopp, L.; Doney, S. C.; Feely, R. A.; Gnanadesikan, A.; Gruber, N.; Ishida, A.; Joos, F.; et al. Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying organisms. *Nature* **2005**, *437*, 681–686.
- (139) Bibby, R.; Cleall-Harding, P.; Rundle, S.; Widdicombe, S.; Spicer, J. Ocean acidification disrupts induced defences in the intertidal gastropod *Littorina littorea*. *Biol. Lett.* **2007**, *3*, 699–701.
- (140) Saderne, V.; Fietzek, P.; Herman, P. M. J. Extreme variations of pCO₂ and pH in a macrophyte meadow of the Baltic Sea in summer: Evidence of the effect of photosynthesis and local upwelling. *PLoS One* **2013**, *8*, e62689.
- (141) Liu, J.; Weinbauer, M. G.; Maier, C.; Dai, M.; Gattuso, J. Effect of ocean acidification on microbial diversity and on microbe-driven biogeochemistry and ecosystem functioning. *Aquat. Microb. Ecol.* **2010**, *61*, 291–305.
- (142) Falkenberg, L. J.; Russell, B. D.; Connell, S. D. Contrasting resource limitations of marine primary producers: Implications for competitive interactions under enriched ${\rm CO_2}$ and nutrient regimes. *Oecologia* **2013**, 172, 575–583.
- (143) Kroeker, K. J.; Micheli, F.; Gambi, M. C. Ocean acidification causes ecosystem shifts via altered competitive interactions. *Nat. Clim. Change* **2013**, *3*, 156–159.
- (144) Fabry, V. J.; Seibel, B. A.; Feely, R. A.; Orr, J. C. Impacts of ocean acidification on marine fauna and ecosystem processes. *ICES J. Mar. Sci.* **2008**, *65*, 414–432.

- (145) Ziveri, P.; de Bernardi, B.; Baumann, K.-H.; Stoll, H. M.; Mortyn, P. G. Sinking of coccolith carbonate and potential contribution to organic carbon ballasting in the deep ocean. *Deep Sea Res.*, *Part II* **2007**, *54*, 659–675.
- (146) Wilson, R. W.; Millero, F. J.; Taylor, J. R.; Walsh, P. J.; Christensen, V.; Jennings, S.; Grosell, M. Contribution of fish to the marine inorganic carbon cycle. *Science* **2009**, 323, 359–362.
- (147) Perry, C. T.; Salter, M. A.; Harborne, A. R.; Crowley, S. F.; Jelks, H. L.; Wilson, R. W. Fish as major carbonate mud producers and missing components of the tropical carbonate factory. *Proc. Natl. Acad. Sci. U.S.A.* **2011**, *108*, 3865–3869.
- (148) Wilson, R.; Gilmour, K.; Henry, R.; Wood, C. Intestinal base excretion in the seawater-adapted rainbow trout: A role in acid-base balance? *J. Exp. Biol.* **1996**, *199*, 2331–2343.
- (149) Wilson, R. W.; Grosell, M. Intestinal bicarbonate secretion in marine teleost fish-source of bicarbonate, pH sensitivity, and consequences for whole animal acid-base and calcium homeostasis. *Biochim. Biophys. Acta* **2003**, *1618*, 163–174.
- (150) Howarth, R.; Chan, F.; Conley, D. J.; Garnier, J.; Doney, S. C.; Marino, R.; Billen, G. Coupled biogeochemical cycles: Eutrophication and hypoxia in temperate estuaries and coastal marine ecosystems. *Front. Ecol. Environ.* **2011**, *9*, 18–26.
- (151) Melzner, F.; Thomsen, J.; Koeve, W.; Oschlies, A.; Gutowska, M. A.; Bange, H. W.; Hansen, H. P.; Körtzinger, A. Future ocean acidification will be amplified by hypoxia in coastal habitats. *Mar Biol.* **2013**, *160*, 1875–1888.
- (152) Diaz, R. J.; Rosenberg, R. Spreading dead zones and consequences for marine ecosystems. *Science* **2008**, 321, 926–929.
- (153) Odum, E. P.; Finn, J. T.; Franz, E. H. Perturbation theory and the subsidy-stress gradient. *BioScience* 1979, 29, 349-352.
- (154) Riebesell, U.; Gattuso, J.-P.; Thingstad, T. F.; Middelburg, J. J. Preface "Arctic ocean acidification: Pelagic ecosystem and biogeochemical responses during a mesocosm study. *Biogeosciences* **2013**, *10*, 5619–5626.
- (155) Zablocki, J. A.; Andersson, A. J.; Bates, N. R. Diel aquatic CO₂ system dynamics of a bermudian mangrove environment. *Aquat. Geochem.* **2011**, *17*, 841–859.
- (156) Falkenberg, L. J.; Russell, B. D.; Connell, S. D. Stability of strong species interactions resist the synergistic effects of local and global pollution in kelp forests. *PLoS One* **2012**, *7*, e33841.